



REVIEW

Genetic counseling for prion disease: Updates and best practices



Jill S. Goldman¹, Sonia M. Vallabh^{2,3,4,5,*} 

¹Columbia University Irving Medical Center, New York, NY; ²Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, Cambridge, MA; ³Henry and Allison McCance Center for Brain Health, Massachusetts General Hospital, Boston, MA; ⁴Department of Neurology, Massachusetts General Hospital and Harvard Medical School, Boston, MA; ⁵Prion Alliance, Cambridge, MA

ARTICLE INFO

Article history:

Received 28 February 2022

Received in revised form

13 June 2022

Accepted 15 June 2022

Available online 12 July 2022

Keywords:

Creutzfeldt-Jakob disease (CJD)

Fatal familial insomnia (FFI)

Gerstmann-Sträussler-Scheinker disease

(GSS)

Prion disease

PRNP

ABSTRACT

Prion disease is a rare, fatal, and often rapidly progressive neurodegenerative disease. Ten to fifteen percent of cases are caused by autosomal dominant gain-of-function variants in the prion protein gene, *PRNP*. Rarity and phenotypic variability complicate diagnosis, often obscuring family history and leaving families unprepared for the genetic implications of an index case. Several recent developments inspire this update in best practices for prion disease genetic counseling. A new prion-detection assay has transformed symptomatic diagnosis. Meanwhile, penetrance, age of onset, and duration of illness have been systematically characterized across *PRNP* variants in a global cohort. Clinically, the traditional genotype–phenotype correlation has weakened over time, and the term genetic prion disease may now better serve providers than the historical subtypes Creutzfeldt-Jakob disease, fatal familial insomnia, and Gerstmann-Sträussler-Scheinker disease. Finally, in the age of genetically targeted therapies, clinical trials for prion disease are being envisaged, and healthy at-risk individuals may be best positioned to benefit. Such individuals need to be able to access clinical services for genetic counseling and testing. Thus, this update on the genetics of prion disease and best practices for genetic counseling for this disease aims to provide the information needed to expand genetic counseling services.

© 2022 The Authors. Published by Elsevier Inc. on behalf of American College of Medical Genetics and Genomics. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

Human prion disease is a rare, fatal neurodegenerative disease, major subtypes of which include Creutzfeldt-Jakob disease (CJD), fatal familial insomnia (FFI), and Gerstmann-Sträussler-Scheinker disease (GSS). Prion disease is the cause of roughly 1 in 6000 deaths,¹ with an incidence of 1 to 2 cases per million population per year.² Although 85%

of cases are sporadic, with no known genetic or environmental trigger, approximately 10% to 15% are genetic, arising from autosomal dominant protein-altering variants in *PRNP*.³ Acquired cases, made famous by the mad cow epidemic, are rare today. Regardless of etiology, prion disease is caused by the misfolding of the prion protein, which is termed PrP or PrP^c in its normal state. Misfolded PrP conformers, termed PrP^{Sc} or prions, act as templates for

*Correspondence and requests for materials should be addressed to Sonia M. Vallabh, Stanley Center for Psychiatric Research, Broad Institute of MIT and Harvard, 75 Ames St., Cambridge, MA 02142. E-mail address: svallabh@broadinstitute.org

doi: <https://doi.org/10.1016/j.gim.2022.06.003>

1098-3600/© 2022 The Authors. Published by Elsevier Inc. on behalf of American College of Medical Genetics and Genomics. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

conformational conversion of additional PrP molecules.⁴ Accumulation of prions in the brain gives rise to neurodegeneration with characteristic pathology; broad spongiform change is pathognomonic, whereas plaque and localized regional pathology are also seen in some cases. Although early symptoms vary widely, most cases rapidly advance into a progressive dementia with average duration of less than half a year.⁵ Variability is seen in both presentation and duration of genetic prion disease cases and shows some association with *PRNP* genotype as further discussed in later sections. Diagnosis of prion disease is often delayed unless the patient is seen by a neurologist familiar with the disease.⁶ Historical diagnostic tools have included selected magnetic resonance imaging, electroencephalogram, and nonspecific fluid biomarkers of neuronal damage.⁷ Brain biopsy, formerly used in diagnosis, is presently discouraged. More recently, the disease-specific real-time quaking induced conversion assay, which detects prion seeds in cerebrospinal fluid or brain tissue, has revolutionized both pre- and postmortem diagnosis, offering >90% sensitivity and specificity, particularly for sporadic forms of prion disease.⁸ The assay's reduced sensitivity to some genetic subtypes can be complemented by targeted sequencing of *PRNP*, which should be routinely offered for all suspected cases of prion disease, whether or not a family history is immediately apparent.³

In this article, we will consider the genetic forms of prion disease. More than half of these cases lack a documented family history and thus are more appropriately referred to as genetic than familial or hereditary.⁹ Absent family history can be due to de novo pathogenic variants, incomplete penetrance, misattributed parenthood, adoption or estrangement, early death by other causes, or misdiagnosis of previous generations. Given the possibility of a genetic etiology, clinicians should consider offering genetic counseling and testing when diagnosing prion disease. Learning of genetic prion disease in the family can be shocking and terrifying, especially because of broader family implications. Families often struggle to find a provider knowledgeable about the disease and equipped to counsel about its genetic risk. This article will address issues pertinent to prion disease genetic counseling, including disease presentation, genetics, psychosocial counseling, and resources available to support patients, families, and those at risk.

Genetic Forms of Prion Disease

Genetic prion disease is caused by protein-altering variants in *PRNP* located on chromosome 20p13. Of variants with strong evidence of pathogenicity, most are missense variants, but octapeptide repeat insertion (OPRI) variants and, in rare cases, truncating and frameshift variants can also cause disease.³ Although many *PRNP* variants reported as pathogenic have subsequently been shown to convey modest or no risk, several variants cause disease with high penetrance, as

shown by enrichment of these variants in prion disease cases over population controls and percentage of cases with positive family history.¹⁰ The most common such highly penetrant variants are E200K, P102L, D178N, 6-OPRI, 5-OPRI, A117V, and P105L,¹⁰ all of which meet American College of Medical Genetics and Genomics/Association for Molecular Pathology (ACMG/AMP) criteria PS4 (enrichment in cases over controls).¹¹ A total of 20 additional variants have been classified as likely highly penetrant on the basis of a combination of ACMG/AMP criteria PM2 (absence from controls in a large population data set) as well as either PS2 (de novo variant reported in a patient with no family history) or PP1 (cosegregation with disease in a pedigree with multiple affected); see [Supplemental Table 1](#) for a full list. A handful of less penetrant variants, including V210I, V180I, and M232R, appear to convey 0.1% to 10% lifetime risk.¹⁰

In addition to these pathogenic variants, *PRNP* harbors 1 common sequence variant at codon 129, which can be occupied by either a methionine (M) or valine (V) residue. Codon 129 genotype influences duration of disease in some genetic prion disease, as well as risk in sporadic and iatrogenic prion disease.³ This codon does not appear to influence age of onset for the most common genotypes in genetic prion disease,¹² although it may in rare cases.

The pathogenic variants listed earlier are associated with a spectrum of prion disease presentations. Some variants, including E200K, D178N-129V, and V210I, have historically been associated with the clinical term genetic CJD. The P102L, P105L and A117V variants were traditionally associated with GSS, whereas the D178N-129M variant was associated with FFI. The terms CJD, GSS, and FFI predate the discovery of *PRNP* as the single causal gene unifying all prion disease, and the field now recognizes abundant phenotypic heterogeneity within and overlap between these historical subtypes.^{3,13,14} The current understanding of the most clinically relevant *PRNP* variants is described later in this article.

Classical phenotypes

Though most cases of prion disease converge toward progressive dementia, akinetic mutism, and ultimately, terminal illness, early symptoms vary widely in a manner not faithfully predicted by genotype. Diverse presentations have been reported not only within variants¹⁵⁻²¹ but even within the same affected family^{13,22,23} and between affected monozygotic twins.²⁴⁻²⁶ However, despite its erosion over time, it is useful to be aware of the classical genotype-phenotype correlation ([Table 1](#)). First, although a simplified categorization can mislead in individual cases, some meaningful if imperfect group-wise distinctions persist, such as rapidly versus slowly progressive variants.²⁷ Second, patient and family awareness of classical subtypes may prime them to expect or notice certain symptoms. Educating families about the phenotypic spectrum may help to resolve confusion in instances when cases have appeared phenotypically mismatched to historical subtype, or have

Table 1 Phenotypes and *PRNP* variants historically associated with the 3 classical subtypes of genetic prion disease

Clinical Term for Historical Prion Disease Subtype	Classical Symptoms Historically Associated With This Subtype	<i>PRNP</i> Variants Historically Associated With This Subtype
Genetic Creutzfeldt-Jakob disease	Prominent early cognitive symptoms: memory decline, dementia; also ataxia, myoclonus, pyramidal and extrapyramidal signs, behavioral change, psychiatric symptoms such as hallucinations, delusions, and depression; rapidly progressive	E200K, D178N-129V, V210I, V180I, M232R
Fatal familial insomnia	Prominent early dysautonomic symptoms: sleep dysregulation, sympathetic overactivity, endocrine abnormalities; also, abnormal gait, weakness, hallucinations, cognitive impairment, dementia; rapidly progressive	D178N-129M
Gerstmann-Sträussler-Scheinker disease	Prominent early motor symptoms: progressive cerebellar ataxia, parkinsonism, muscle weakness; also, cognitive impairment, sensory defects, behavioral change, dementia; slowly progressive.	P102L, A117V, P105L, F198S

Symptoms summarized from Takada et al.²⁷ and Takada et al.¹⁴ This list includes both high-penetrance pathogenic variants and risk factors with low to modest penetrance.¹⁰

presented differently within a family despite shared genotype.

Some pathogenic *PRNP* variants have historically eluded the framework mentioned earlier, including the OPRI variants. *PRNP* normally has a region of five 24-base pair octapeptide repeats. Although both deletions and insertions can occur, Mendelian segregation is best established for insertions of 5 to 12 repeats. Clinical progression varies widely and is imperfectly predicted by repeat length. Both CJD-like and GSS-like presentations have been reported within a given OPRI family, and mixed CJD-like and GSS-like pathology have been reported even within the same brain, offering further challenge to the diagnostic boundaries between these subtypes.²⁷

Rare nonsense variants can also cause prion disease by producing a truncated version of the protein that fails to properly localize to the plasma membrane. These variants tend to present with a longer disease course and highly variable range of phenotypes atypical of prion disease. Some cases are mistaken for Alzheimer disease or behavioral variant frontotemporal dementia owing to their gradual course variably defined by behavioral, cognitive, and motor decline.^{14,27} By contrast, a subset of these variants, including R163X, cause a yet more unusual phenotype including sensorimotor neuropathy, chronic diarrhea, and urinary dysfunction, with dementia emerging only late in the average 20-year course. The prominence of peripheral symptoms may raise suspicion of a hereditary sensory and autonomic neuropathy or transthyretin familial amyloid polyneuropathy.²⁸ This wide variability in the first symptoms of genetic prion disease highlights the critical importance of performing *PRNP* sequencing at the earliest opportunity.

Penetrance in genetic prion disease

As penetrance estimates for *PRNP* variants are continuously subject to update, clinicians should approach the prion disease literature with caution. Some reported variants

appear to have low penetrance or be benign despite reports to the contrary in PubMed. Case reports must be approached with the caveat that a variant seen only once or a few times may be a benign bystander in a sporadic case; see Mok et al.²⁹ for an example of how to evaluate such cases consistent with ACMG/AMP guidance. Confidence in pathogenicity can be gained from evidence of Mendelian segregation and/or a convincing enrichment of the variant in cases compared with population controls.^{10,12} According to recent studies using these methods, several variants (Figure 1) have evidence of high penetrance, conveying lifetime risk of >90%.^{10,12} The 3 most common highly penetrant genetic prion disease variants—E200K, D178N, and P102L—collectively account for 53% of all genetic prion disease cases, and 85% of those caused by a highly penetrant variant.¹⁰

Certain *PRNP* variants appear to be low-penetrance risk factors, based both on prevalence in cases vs controls and frequency of family history.¹⁰ V210I is the most common such variant, conveying an estimated 10% lifetime risk. The variants V180I and M232R correspond to an estimated 1% and 0.1% lifetime risk, respectively.

Although future analysis of larger case and control data sets may refine penetrance estimates, the overall categorization of the aforementioned variants as high or low penetrance is unlikely to change. More data may, however, enable interpretation of variants seen too rarely in either prion disease cases or the general population to enable a meaningful assignment of pathogenicity thus far. Supplemental Table 1 provides an overview of *PRNP* variants reported to date, including very rare variants, and evidence available at present to support high penetrance and/or increased risk.

Age of onset in genetic prion disease

Age of onset varies dramatically for all genetic prion disease variants (Table 2), and no factors, including sex or parental

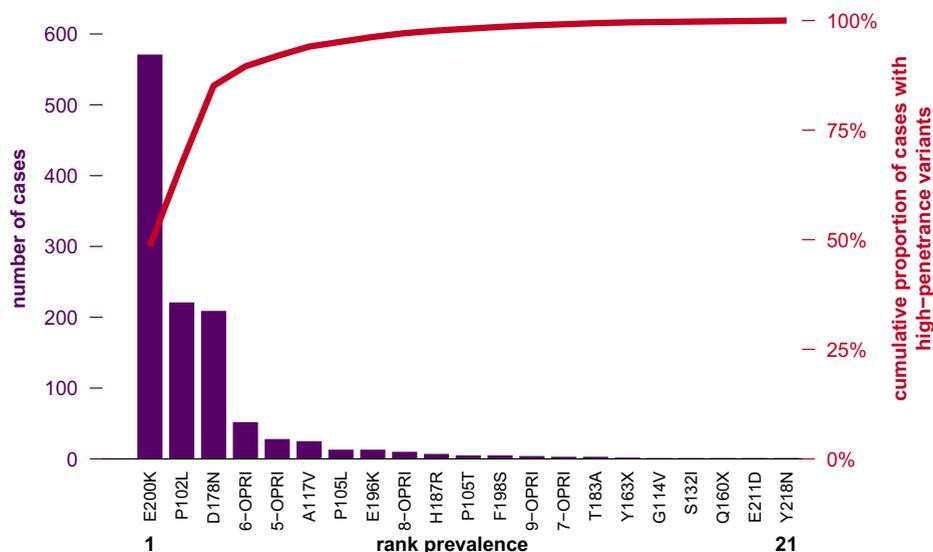


Figure 1 Prevalence and relative contribution to overall case load of highly penetrant *PRNP* variants. Genetic variants with evidence for high penetrance¹² are plotted according to the number of corresponding cases in a case series of 10,460 genetic prion disease patients gathered from prion disease centers in 9 countries. The cumulative proportion of high penetrance cases accounted for by these variants is shown on the right axis. Figure reproduced with permission from https://github.com/ericminikel/prnp_onset/blob/master/figures/figure_s1.pdf, see Minikel et al¹² for details. OPRI, octapeptide repeat insertion.

age of onset, are known to predict it. In families with extensive family history, age of onset variability may be a source of confusion. Families may theorize triggers for symptomatic onset, including a recent stressful incident or medical event. However, there is no evidence to support an environmental, behavioral, or medical trigger for onset of genetic prion disease.

Unlike in Huntington disease, an individual's age of onset does not appear to be predicted by their affected parent's age of onset,¹² including that there is a lack of anticipation or systematic trend toward earlier onset in subsequent generations.³⁰ Unless raised by the family, introducing the concept of anticipation, even to explain that it is inapplicable, is likely to cause only undue distress.

Rate of progression in genetic prion disease

Notwithstanding significant within-group variation, genetic prion disease variants can be categorized as rapidly progressive (typically progressing from first symptoms to death in less than 3 years) or slowly progressive (total disease duration averaging 3 or more years).²⁷ Roughly two-thirds of genetic prion disease cases are rapidly progressive, including those caused by the relatively common E200K and D178N variants;¹⁰ 50% of these patients die within 1 year of first symptoms and only a small minority survive to 2 years after onset (Figure 2). By contrast, OPRI variants and those traditionally linked to GSS tend toward a longer disease course. However greater variability in duration for these variants precludes meaningful individual-level predictions. P102L, the most common slowly progressive variant, can correspond to a disease course of 1 year or 15 years. Notably these data are confounded by end-of-life

decisions and do not capture when a patient became dependent on intubation or ventilation, which can dramatically extend the nominal course of prion disease.³¹

Founder populations

Some genetic prion disease variants show geographical clustering due to founder effects. The variants V210I and V180I risk factors are more common among those of Italian and Japanese ancestry, respectively, whereas the E200K variant is unusually prevalent among Libyan Jews in Israel and Slovaks.²⁷ Despite this enrichment, the E200K, D178N, and P102L variants appear to have arisen independently in many populations; all 3 are found around the world.¹⁰

Genetic Counseling for Prion Disease

Preparing for counseling on prion disease

Despite its rarity, prion disease is well-characterized and resources exist to aid in preparation for counseling sessions. Although care teams must commit to thorough preparation, lack of experience with prion disease per se should not be considered a barrier to providing counseling. Many families must conduct an extensive search to find a provider willing to provide counseling and testing. This article aims to fill this gap by providing the best practices for meeting the needs of these families.

Ideally a multidisciplinary team, including a neurologist, a genetic counselor, and a social worker will have the opportunity to manage genetic prion disease cases together. Key preparation will include understanding the penetrance

Table 2 Age of onset for the 7 most common highly penetrant genetic prion disease variants, ranked by number of cases documented in a case series of 1094 genetic prion disease cases gathered from 9 international prion disease centers

Pathogenic Variant	Without Censored Data		Survival Curve Including Censored Data		
	Mean \pm SD	N	Median (IQR)	Range	N
E200K	61.3 \pm 10.0	456	62 (55-68)	31-92	506
D178N	51.3 \pm 11.8	256	53 (46-60)	12-89 ^a	289
P102L	53.7 \pm 10.6	193	56 (47-60)	22-75	206
6-OPRI	35.1 \pm 5.8	31	35 (32-39)	23-47	34
A117V	41.2 \pm 7.8	26	41 (37-45)	25-58	28
5-OPRI	46.8 \pm 6.0	14	49 (44-53)	34-56	18
P105L	46.5 \pm 8.5	13	47 (40-51)	31-61	13

Data reproduced with permission from https://github.com/ericminikel/prnp_onset/blob/master/figures/table_1_table_s3.xls, see Minikel et al¹² for details. Censored data refers to individuals who had not experienced onset at last observation.

IQR, interquartile range; OPRI, octapeptide repeat insertion.

^aDenotes a pathogenic variant-positive individual still healthy at age 89.

of the relevant variant, if known, as well the wide range in age of onset and phenotypic presentation associated with all variants. Both families of prion disease patients undergoing *PRNP* sequencing and at-risk individuals considering predictive testing should have before and after test genetic counseling.

Testing

The team should be familiar with testing protocols in prion disease. Because of its technically transmissible nature, most developed countries have a national surveillance center that performs centralized testing for prion disease. In the United States, the National Prion Disease Pathology Surveillance Center in Cleveland, Ohio, routinely handles diagnostic testing, including cerebrospinal fluid real-time quaking induced conversion assay for premortem diagnosis, as well as postmortem autopsy. It also performs targeted *PRNP* sequencing on both symptomatic patients and those at risk. At the time of preparing this manuscript, predictive testing was free of charge for those with a first-degree relative confirmed to have prion disease. Results are returned to an ordering health care provider to be shared with the patient or family. In addition to providing testing, the Surveillance Center serves as a resource and biobank for the prion disease research field and is equipped to receive postmortem tissue should a patient's family wish to donate tissue to research. Although several other clinical laboratories also offer *PRNP* genetic testing either as a single gene test or part of a panel, ordering providers must be mindful of testing techniques and coverage, as OPRI are not typically detected by next-generation sequencing (NGS). Therefore, unless there is a known *PRNP* point variant in the family, a protocol designed to detect OPRI, such as gel electrophoresis,³² allele-specific Sanger sequencing, or long-read sequencing³³ should be employed. Bearing in mind this

limitation of NGS, in the event that prion disease is part of a broad differential diagnosis of a neurodegenerative disease and there is a family history of a similar condition, a large NGS dementia panel may be considered as a first step to rule out conditions such as familial Alzheimer's disease or frontotemporal dementia.

Transmission concerns

If raised by the patient or family, providers should be prepared to speak to questions about the horizontally transmissible nature of prion disease. It is crucial to reassure concerned families that misfolded PrP is transmitted from person to person only through extraordinary circumstances such as brain-to-brain contact in the context of a medical procedure.³⁴ Prion disease is not acquired through any normal activity, including sharing a household, intimate contact, caretaking activities, or routine medical or dental care. In addition, available evidence suggests that individuals with pathogenic *PRNP* variants are healthy for the vast majority of their lives, with prions appearing in the central nervous system only briefly before onset of clinical symptoms.³⁵ In genetic prion disease families, the disease tracks strictly with the inheritance of the pathogenic variant; there is no evidence to suggest that prions are passed vertically through breastfeeding or pregnancy.³⁶

Many blood banks maintain cautious policies regarding prion disease risk and individuals with either a family history of prion disease or known pathogenic *PRNP* variant may be distressed to find themselves ineligible to donate blood. There have been 4 cases of human-to-human transmission of the peripherally acquired prion disease subtype variant CJD after blood transfusion.³⁷ In the context of genetic prion disease, such prohibitions reflect an abundance of caution rather than an established risk. Blood-based transmission of prion disease has not been reported for any form of human prion disease other than acquired variant CJD, and even at the symptomatic stage of disease, blood is considered a "no detectable infectivity" tissue by the World Health Organization.³⁸ Risk should be particularly low in asymptomatic individuals at risk for genetic prion disease given that prions are generally not detectable even in their spinal fluid.³⁵ Therefore, individuals from genetic prion disease families should be reassured that their blood poses no known transmission risk.

Resources

At present, no disease-modifying treatment for prion disease is available. It will be important to convey this information in balance with a high-level overview of the state of the prion disease field. Like many rare diseases, prion disease is profoundly isolating. Families may mistakenly assume that no one else can relate to their plight or that such a rare disease lacks prospects for scientific progress. It is critical to emphasize that despite the rarity of prion disease, the

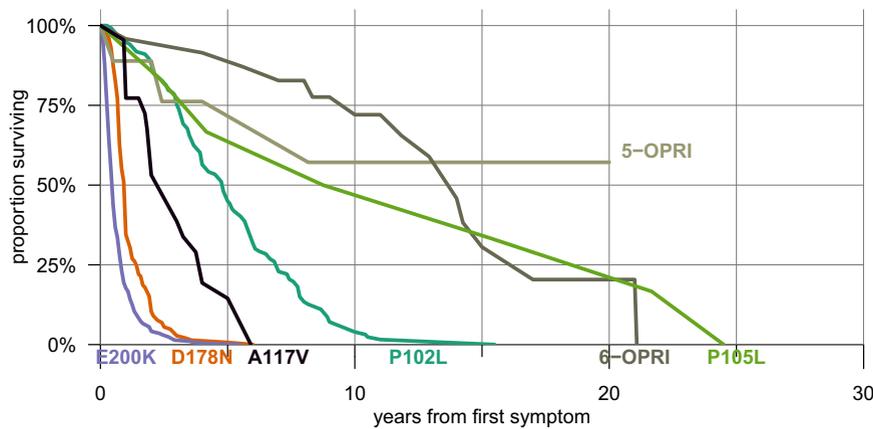


Figure 2 Rate of progression of genetic prion disease for the 7 most common highly penetrant *PRNP* variants. Figure reproduced with permission from https://github.com/ericminikel/prnp_onset/blob/master/figures/figure_s2.pdf, see Minikel et al¹² for details. OPRI, octapeptide repeat insertion.

community of affected patients and families is strikingly well-organized and the disease itself is well-understood by a large and active research community. Genetic counselors and clinicians can refer patients, families, and at-risk individuals to resources available to help them feel less alone, track down answers, and understand the progress being made toward meaningful therapies (Table 3).

Diagnostic genetic counseling session

In many cases, a genetic counseling session may be triggered by a symptomatic case within the family and may occur either before or after the patient has been tested. Given the disease's rapid progression, family members are likely to attend without the proband. Because so many cases lack a family history, families may be referred for genetic counseling without any prior knowledge of either the disease or the attendant genetic risk. By contrast, some families may arrive already expert on a disease that they have witnessed many times.

As with any genetic counseling, sessions should start by determining why the client was motivated to schedule a consultation and what they wish to achieve. Understanding the client's disease experience is essential for providing both the right information and the needed compassion. Families taxed by a long diagnostic odyssey and by the rapidity of their loved one's decline may arrive depleted and learning that the disease could be genetic can come as a shock. Unless genetic counselors take the time to allow their clients to grieve before diving into the genetics of the disease, little will be heard or retained.³⁹

When the clients are ready, a detailed pedigree should be taken noting cause and age of death, disease duration, and neurologic and psychiatric history in at least 3 generations. Targeted questions should focus on cognitive impairment, movement disorders (eg, parkinsonism, ataxia), seizures, insomnia, depression, anxiety, personality change, and psychosis. Once obtained, the pedigree can be used to personalize the educational part of the session.

Care should be taken to tailor background explanation to the baseline scientific and medical literacy of the clients, as well as to their specific understanding of prion disease. With that caveat, the informational part of the session should summarize the following:

- Definition of prion, types of prion disease (sporadic, genetic), the pathogenesis of prion disease
- Basic genetics: autosomal dominant inheritance, structure of DNA, variants
- The concept of de novo pathogenic variants, and other causes of missing family history
- Inter- and intra-familial phenotypic variability (presentation, age of onset, duration of disease). Referring to the pedigree to show variability can be helpful.
- If known, specific information about the family variant, especially prognosis and penetrance. Referring to the pedigree to show reduced penetrance and missing family history can aid understanding.
- As individuals try to understand their own risk or that of family members, care should be taken to distinguish between the following:
 - A priori, there is a 10% to 15% chance that a prion disease case is genetic.
 - If the case is confirmed to be genetic, there is a 50% chance that a child of an affected parent inherited the causal variant.
 - If inherited, risk may still be influenced by that variant's penetrance.

After the informational portion of the session, the genetic counselor should assess the family's understanding of the genetic risk. Specific issues to be explored are risk to family members present, sharing of information with non-attending family members, and whether any family members might be interested in predictive testing. The counselor can offer to talk with other family members or write an informational family letter.

Table 3 Resources to support patients and families affected by prion disease

Organization	Mission	Key Activities
CJD Foundation (cjd.foundation.org)	A US-based nonprofit organization with the mission of supporting families affected by prion disease, raising awareness, and supporting prion disease-related medical education and research	The CJD Foundation operates a patient and family helpline, provides educational materials to both families and medical professionals, hosts regular workshops and teleconferences, advocates for prion disease research and surveillance funding, makes research grants, and convenes an annual meeting that brings together affected families and prion disease experts
CJD International Support Alliance	International network of nonprofit organizations committed to improving the lives of prion disease patients and families around the world.	The CJD ISA serves as an umbrella organization for patient support groups around the world, supports patients to connect with care and resources in their country, and supports those interested in starting new patient groups in countries not yet served
Prion Alliance (prionalliance.org)	A US-based non-profit devoted to funding scientific research toward a treatment for prion disease.	Prion Alliance, which is run by patient-scientists personally affected by genetic prion disease, fundraises to support the development of prion disease therapeutics, in addition to maintaining a frequently updated prion disease FAQ; an associated scientific blog, cureffi.org , reviews key topics in prion disease and adjacent biomedical domains, for providers or patients/families interested in engaging prion science in more detail.
Prion Registry (PrionRegistry.org)	A private, secure, global online registry for those affected by prion disease in any way (patients, at-risk individuals whether tested or untested, family and friends who wish to register as controls).	Co-led by the Prion Alliance, CJD Foundation, and CJD ISA, the Prion Registry connects the prion disease community with opportunities to participate in research studies or future clinical trials. Although intake data from each participant remains strictly private, anonymized group-level data from the registry are available to qualified researchers to facilitate therapeutic research and future trials.

CJD, Creutzfeldt-Jakob disease; *FAQ*, frequently asked questions, *ISA*, International Support Alliance.

If genetic testing on the proband has not yet been performed, the possible types of results (positive, negative, variant of uncertain significance) should be explained, as well as the implications for the family. Finally, the genetic counselor should lead the family through a discussion on whether a genetic test on the affected person is in the best interest of that person and the family. Because families may be overwhelmed by having to manage the affected person's needs, alternatives to testing during the lifetime should be offered, including banking DNA and testing future autopsy tissue. If appropriate, an informed consent should be signed. Families should also be provided with the resources listed at the top of this section. The CJD Foundation, in particular, is well-equipped to support those caring for a currently symptomatic loved one.

Predictive genetic counseling for prion disease

In the context of predictive testing, it is important to recognize the wide spectrum of preference present in the at-risk community. The appropriate path for any at-risk individual will need to be determined on a case-by-case basis.

Because prion disease is a fatal neurodegenerative disease, providers should be prepared to follow a modified version of the Huntington disease protocol. In weighing the appropriate level of procedure and caution, genetic counselors will need to listen to the client's needs and

preferences, whereas also being sensitive to the factors that are correlated with rare but concerning adverse outcomes after predictive testing: a prior history of significant anxiety or depression, lack of planning for the future, strong identification with the disease that is not adaptable to an unexpected result, a poor support system, or poor communication with family.⁴⁰⁻⁴⁷ Notably, negative reactions can accompany either a positive or negative result.^{43,48} For individuals with either identified risk factors or significant uncertainties about their readiness for testing, access to an evaluation by a psychiatrist familiar with predictive genetic testing for neurodegenerative disease may be helpful.⁴⁹⁻⁵¹ If access to psychiatry is limited, genetic counselors may involve another clinician to evaluate the client's readiness. In such cases it may also be reasonable to recommend a minimum 1-month interval between genetic counseling and genetic testing, to allow the client time to process new information and possibly change their mind.⁵¹

Despite the concern that they generate, adverse reactions to testing are uncommon. The literature consistently shows that most people who undergo predictive testing for a neurodegenerative disease adapt to their results without long-term consequence.^{47,51-54} Although some studies report after-test divergence in distress levels between those who test positive and negative, these gaps typically close

within weeks to months.^{42,46,55,56} Many people report important benefits from receiving their genetic information, regardless of outcome.^{47,57} Relief from uncertainty can be a significant psychological factor, capable of more than compensating for an unwanted result^{42,57,58} and the act of gathering information through testing can itself be an effective coping mechanism.⁵⁹ A recent study in genetic prion disease echoes the broader finding that although having a genetic disease in the family is stressful, predictive testing does not increase this stress level above baseline.⁶⁰ There is some indication that predictive testing rates are gradually rising in genetic prion disease,⁶¹ in keeping with increased rates of genetic testing in neurology clinics in general^{62,63} and perhaps reflective of a greater openness to predictive testing among young adults.⁶⁴

Just as some individuals will approach testing with ambivalence, others will approach it with self-knowledge that they wish to access their genetic information—whether to relieve unwanted ambiguity, assert personal control, begin planning for the future, or simply out of a “need to know.” The testing protocol should be flexible for clients who understand the disease, lack risk factors for bad outcomes, and are confident that they wish to test. It is important to recognize that whereas some clients may feel pressured by family members or physicians to pursue genetic testing, others may feel pressure to not pursue genetic testing. Regardless, this decision is ultimately the individual’s alone.⁵⁰ A genetic counselor should be prepared to serve as a buffer against either form of pressure, on a case-by-case basis. For clients with high-risk characteristics, it may be appropriate for genetic counselors to slow the process down until the individual is adequately informed. For others, the genetic counselor may serve the equally critical role of helping to minimize barriers and delays that may themselves become acute sources of stress at an already demanding time.

Predictive genetic testing counseling sessions

Among individuals seeking predictive testing, awareness of family history will vary from multigenerational knowledge to recent first encounters with the disease. Thus, as with diagnostic testing, it is essential to ask about the client’s experience with the disease, validate their feelings, and explore their motivation for genetic counseling.

The informational part of the session mirrors a diagnostic counseling session, with special emphasis on the relevant variant’s phenotypic variation and the penetrance. It is important for the client to understand that if they test positive, their age of onset and presentation could differ from other family members. The discussion should encompass the Genetic Information Non-discrimination Act (GINA,) life/long-term care insurance, and reproductive options. It can be noted that in vitro fertilization with preimplantation genetic testing has been used successfully in genetic prion disease.⁶⁵ Creative options around disclosure of results should also be discussed, including nondisclosure in vitro

fertilization with preimplantation genetic testing (IVF-PGT) and the possibility of depositing DNA or holding a genetic test result in reserve for future use by the client or their family.⁵⁰

In addition to the aforementioned points, clients should be given access to the resources highlighted at the top of this section, and should be introduced to the presence of a well-organized support network and active research community in prion disease. With the preface that participation is always an individual’s choice that can be undertaken or revoked at any time, they should be informed that there are opportunities for healthy individuals at risk for prion disease to participate in research. Some clients will feel powerfully motivated to contribute to scientific progress against this disease^{57,61} and will be empowered by the knowledge that in a rare disease, every participant’s contribution is significant. It is also important to share with clients that as in many brain diseases, research indicates that effective prion disease treatments, once available, will have the greatest impact if given before symptoms arise.^{66,67} This understanding has generated momentum for preventive clinical trials in healthy individuals at known risk for genetic prion disease.⁶⁶

The psychosocial portion of the session should be considered anticipatory guidance and is key to helping the client make an informed decision about whether to test. In the spirit of transparency, the client should be informed that most people adapt well to their results regardless of outcome and that factors associated with adverse outcomes include prior significant depression or anxiety, inflexible assumptions about results, and inadequate social support.

The following topics and questions should then be explored:

- How do you imagine a positive or negative result would affect you emotionally over the short-term? Over the long-term?
- In what ways would a positive or negative result change your choices and quality of life?
- How would a positive result affect your significant other, your parents, your siblings, or other key people in your life?
- Do you think it would be useful to you to have a family discussion before testing, to discuss who would want to know your results, and how you would support each other if results differed?
- Would you consider seeing a therapist for emotional support? Do you believe this would be useful for you?

Difficult questions may arise in the discussion of whether results will be communicated to family members. Although there may be an ethical duty to warn, not all individuals will want to do so. Many may be concerned about raising anxiety, particularly in their children. The ideal way of dealing with this issue is for the person being tested to have a discussion with family members before receiving results. They can frame the discussion hypothetically and ask whether the

family member would want to know the result if the at-risk person chose to test. This gives the family members the right not to know and clarifies who to tell. Parents who are hesitant to tell adult children should be forewarned that withholding this information can create unwanted situations such as a pregnancy. Disclosing results to adult children may cause anxiety but gives them the option to pursue their own predictive testing.

Clients are strongly advised to bring a support person with them to both their genetic counseling and result sessions. If present, the following questions should be explored with the support person:

- How do you imagine _____ would respond to a positive or negative test result over the short-term? Over the long-term?
- Are you worried about how they will cope?
- Would you consider seeing a therapist for emotional support? Do you believe this would be useful for you?

These topics often produce emotional responses from the client and from the support person. The genetic counselor should validate their feelings and can also mirror and reinforce signs of strength, resilience and proactiveness that emerge in discussion.³⁹ They may also need to challenge misconceptions. If serious concerns arise, including signs of significant anxiety or depression or discrepant views that could influence the outcome of testing, the genetic counselor can raise for discussion the options of deferring testing, or providing referrals for individual or couples counseling.

Result sessions

It is highly recommended that at least 1 support person attend the result session. If a client believes it will be a net source of support, result sessions may include multiple family members who can support each other and plan together whether and how to communicate results to other family members. The genetic counselor can offer a family meeting or family letter to aid communication. The genetic counselor conveying results should be empathetic yet straightforward. Time should be given to allow the client to process the results. The counselor may offer to leave the room for a short time. The counselor should offer to answer all questions and explore feelings. The genetic counselor can suggest that an appointment be made with a therapist soon after the session. However, this is a recommendation, not a requirement.⁵⁰

Additional questions about horizontal transmission may emerge at this (or any) session. As discussed earlier, concerned individuals should be reassured that person-to-person transmission of misfolded PrP occurs only under exceptional circumstances and that even at the symptomatic stage, prion disease patients pose no hazard to their loved ones.

A follow-up phone call or in person appointment should be offered the week after the result disclosure to answer questions and address concerns.

Conclusion

Prion disease is devastating for both patients and families. Often a long search for a diagnosis ends with a family receiving the news that their loved one has a rapidly progressive fatal disease. This situation is made worse by either knowing that the case fits the pattern of the family disease or by learning for the first time that the disease could be genetic. Genetic counselors are in the unique position of supporting these families in understanding the disease and in making informed decisions about pursuing a genetic diagnosis. Ideally a multidisciplinary team will work together to understand and support client preferences, identify and minimize sources of distress, share relevant resources, and provide, if appropriate, referrals to mental health professionals. For some at-risk individuals, predictive testing may compound the fear and loss of control already set in motion by losing a loved one to prion disease. For others, it may represent one of the few concrete actions available to combat uncertainty, seize agency, and move forward with their lives. At a difficult crossroads, genetic counselors have a rare opportunity to empower clients and families by providing information, respecting autonomy, and sharing opportunities for proactivity, participation, and connection.

Acknowledgments

The authors thank Debbie Yobs and Eric Vallabh Minikel for their helpful comments on this manuscript. SMV was supported by the National Institutes of Health, R01 NS125255.

Author Information

Conceptualization: J.S.G., S.M.V.; Writing—original draft: J.S.G., S.M.V.; Writing—review and editing: J.S.G., S.M.V.

Conflict of Interest

SMV has received speaking fees from Ultragenyx, Illumina, and Biogen, consulting fees from Invitae, and research support in the form of unrestricted charitable contributions from Ionis Pharmaceuticals.

Additional Information

The online version of this article (<https://doi.org/10.1016/j.gim.2022.06.003>) contains supplementary material, which is available to authorized users.

References

- Maddox RA, Person MK, Blevins JE, et al. Prion disease incidence in the United States: 2003-2015. *Neurology*. 2020;94(2):e153–e157. <http://doi.org/10.1212/WNL.0000000000008680>.
- Klug GJ, Wand H, Simpson M, et al. Intensity of human prion disease surveillance predicts observed disease incidence. *J Neurol Neurosurg Psychiatry*. 2013;84(12):1372–1377. <http://doi.org/10.1136/jnnp-2012-304820>.
- Mead S, Lloyd S, Collinge J. Genetic factors in mammalian prion diseases. *Annu Rev Genet*. 2019;53:117–147. <http://doi.org/10.1146/annurev-genet-120213-092352>.
- Prusiner SB. Prions. *Proc Natl Acad Sci U S A*. 1998;95(23):13363–13383. <http://doi.org/10.1073/pnas.95.23.13363>.
- Pocchiarini M, Puopolo M, Croes EA, et al. Predictors of survival in sporadic Creutzfeldt-Jakob disease and other human transmissible spongiform encephalopathies. *Brain*. 2004;127(Pt 10):2348–2359. <http://doi.org/10.1093/brain/awh249>.
- Appleby BS, Rincon-Beardsley TD, Appleby KK, Crain BJ, Wallin MT. Initial diagnoses of patients ultimately diagnosed with prion disease. *J Alzheimers Dis*. 2014;42(3):833–839. <http://doi.org/10.3233/JAD-132465>.
- Figgie MP, Appleby BS. Clinical use of improved diagnostic testing for detection of prion disease. *Viruses*. 2021;13(5):789. <http://doi.org/10.3390/v13050789>.
- Rhoads DD, Wrona A, Foutz A, et al. Diagnosis of prion diseases by RT-QuIC results in improved surveillance. *Neurology*. 2020;95(8):e1017–e1026. <http://doi.org/10.1212/WNL.0000000000010086>.
- Schmitz M, Dittmar K, Llorens F, et al. Hereditary human prion diseases: an update. *Mol Neurobiol*. 2017;54(6):4138–4149. <http://doi.org/10.1007/s12035-016-9918-y>.
- Minikel EV, Vallabh SM, Lek M, et al. Quantifying prion disease penetrance using large population control cohorts. *Sci Transl Med*. 2016;8(322):322ra9. <http://doi.org/10.1126/scitranslmed.aad5169>.
- Richards S, Aziz N, Bale S, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015;17(5):405–424. <http://doi.org/10.1038/gim.2015.30>.
- Minikel EV, Vallabh SM, Orseth MC, et al. Age of onset in genetic prion disease and the design of preventive clinical trials. *Neurology*. 2019;93(2):e125–e134. <http://doi.org/10.1212/WNL.0000000000007745>.
- Collins S, McLean CA, Masters CL. Gerstmann-Sträussler-Scheinker syndrome, fatal familial insomnia, and kuru: a review of these less common human transmissible spongiform encephalopathies. *J Clin Neurosci*. 2001;8(5):387–397. <http://doi.org/10.1054/jocn.2001.0919>.
- Takada LT, Kim MO, Metcalf S, Gala II, Geschwind MD. Prion disease. In: Geschwind DH, Paulson HL, Klein C, eds. *Handbook of Clinical Neurology* [Internet]. Elsevier; 2018:441–464.
- Tesar A, Matej R, Kukul J, et al. Clinical variability in P102L Gerstmann-Sträussler-Scheinker syndrome. *Ann Neurol*. 2019;86(5):643–652. <http://doi.org/10.1002/ana.25579>.
- Zerr I, Giese A, Windl O, et al. Phenotypic variability in fatal familial insomnia (D178N-129M) genotype. *Neurology*. 1998;51(5):1398–1405. <http://doi.org/10.1212/wnl.51.5.1398>.
- Zarranz JJ, Digen A, Atarés B, et al. Phenotypic variability in familial prion diseases due to the D178N mutation. *J Neurol Neurosurg Psychiatry*. 2005;76(11):1491–1496. <http://doi.org/10.1136/jnnp.2004.056606>.
- Sun L, Li X, Lin X, Yan F, Chen K, Xiao S. Familial fatal insomnia with atypical clinical features in a patient with D178N mutation and homozygosity for Met at codon 129 of the prion protein gene. *Prion*. 2015;9(3):228–235. <http://doi.org/10.1080/19336896.2015.1054601>.
- Guerreiro RJ, Vaskov T, Crews C, Singleton A, Hardy J. A case of dementia with PRNP D178Ncis-129M and no insomnia. *Alzheimer Dis Assoc Disord*. 2009;23(4):415–417. <http://doi.org/10.1097/WAD.0b013e3181ae3a76>.
- Taniwaki Y, Hara H, Doh-Ura K, et al. Familial Creutzfeldt-Jakob disease with D178N-129M mutation of PRNP presenting as cerebellar ataxia without insomnia. *J Neurol Neurosurg Psychiatry*. 2000;68(3):388. <http://doi.org/10.1136/jnnp.68.3.388>.
- Fukuoka T, Nakazato Y, Yamamoto M, Miyake A, Mitsufuji T, Yamamoto T. Fatal familial insomnia initially developing parkinsonism mimicking dementia with Lewy bodies. *Intern Med*. 2018;57(18):2719–2722. <http://doi.org/10.2169/internalmedicine.0573-17>.
- McLean CA, Storey E, Gardner RJ, Tannenber AE, Cervenáková L, Brown P. The D178N (cis-129M) “fatal familial insomnia” mutation associated with diverse clinicopathologic phenotypes in an Australian kindred. *Neurology*. 1997;49(2):552–558. <http://doi.org/10.1212/wnl.49.2.552>.
- Synofzik M, Bauer P, Schöls L. Prion mutation D178N with highly variable disease onset and phenotype. *J Neurol Neurosurg Psychiatry*. 2009;80(3):345–346. <http://doi.org/10.1136/jnnp.2008.149922>.
- Honda H, Sasaki K, Takashima H, et al. Different complicated brain pathologies in monozygotic twins with Gerstmann-Sträussler-Scheinker disease. *J Neuropathol Exp Neurol*. 2017;76(10):854–863. <http://doi.org/10.1093/jnen/nlx068>.
- Hamasaki S, Shirabe S, Tsuda R, Yoshimura T, Nakamura T, Eguchi K. Discordant Gerstmann-Sträussler-Scheinker disease in monozygotic twins. *Lancet*. 1998;352(9137):1358–1359. [http://doi.org/10.1016/S0140-6736\(05\)60749-0](http://doi.org/10.1016/S0140-6736(05)60749-0).
- Webb T, Mead S, Beck J, et al. Seven-year discordance in age at onset in monozygotic twins with inherited prion disease (P102L). *Neuropathol Appl Neurobiol*. 2009;35(4):427–432. <http://doi.org/10.1111/j.1365-2990.2009.01012.x>.
- Takada LT, Kim MO, Cleveland RW, et al. Genetic prion disease: experience of a rapidly progressive dementia center in the United States and a review of the literature. *Am J Med Genet B Neuropsychiatr Genet*. 2017;174(1):36–69. <http://doi.org/10.1002/ajmg.b.32505>.
- Mead S, Reilly MM. A new prion disease: relationship with central and peripheral amyloidosis. *Nat Rev Neurol*. 2015;11(2):90–97. <http://doi.org/10.1038/nrneurol.2014.263>.
- Mok TH, Koriath C, Jaunmuktane Z, et al. Evaluating the causality of novel sequence variants in the prion protein gene by example. *Neurobiol Aging*. 2018;71:265.e1–265.e7. <http://doi.org/10.1016/j.neurobiolaging.2018.05.011>.
- Minikel EV, Zerr I, Collins SJ, et al. Ascertainment bias causes false signal of anticipation in genetic prion disease. *Am J Hum Genet*. 2014;95(4):371–382. <http://doi.org/10.1016/j.ajhg.2014.09.003>.
- Nagoshi K, Sadakane A, Nakamura Y, Yamada M, Mizusawa H. Duration of prion disease is longer in Japan than in other countries. *J Epidemiol*. 2011;21:255–262.
- Capellari S, Vital C, Parchi P, et al. Familial prion disease with a novel 144-bp insertion in the prion protein gene in a Basque family. *Neurology*. 1997;49(1):133–141. <http://doi.org/10.1212/wnl.49.1.133>.
- Kroll F, Dimitriadis A, Campbell T, et al. Prion protein gene mutation detection using long-read nanopore sequencing. *Sci Rep*. 2022;12:8284. <http://doi.org/10.1101/2022.03.06.22271294>.
- Will RG. Acquired prion disease: iatrogenic CJD, variant CJD, kuru. *Br Med Bull*. 2003;66(1):255–265. <http://doi.org/10.1093/bmb/66.1.255>.
- Vallabh SM, Minikel EV, Williams VJ, et al. Cerebrospinal fluid and plasma biomarkers in individuals at risk for genetic prion disease. *BMC Med*. 2020;18(1):140. <http://doi.org/10.1186/s12916-020-01608-8>.
- Luk CC, Mathiason CK, Orrù CD, et al. Creutzfeldt-Jakob disease in pregnancy: the use of modified RT-QuIC to determine infectivity in placental tissues. *Prion*. 2021;15(1):107–111. <http://doi.org/10.1080/19336896.2021.1933872>.
- Urwin PJM, Mackenzie JM, Llewelyn CA, Will RG, Hewitt PE. Creutzfeldt-Jakob disease and blood transfusion: updated results of the UK Transfusion Medicine Epidemiology Review Study. *Vox Sang*. 2016;110(4):310–316. <http://doi.org/10.1111/vox.12371>.
- World Health Organization. WHO manual for surveillance of human transmissible spongiform encephalopathies including variant Creutzfeldt-Jakob disease. World Health Organization; 2003. Accessed July 1, 2022. <https://apps.who.int/iris/handle/10665/42656>.

39. MacLeod R, Metcalfe A, Ferrer-Duch M. A family systems approach to genetic counseling: development of narrative interventions. *J Genet Couns.* 2021;30(1):22–29. <http://doi.org/10.1002/jgc4.1377>.
40. Almqvist EW, Bloch M, Brinkman R, Craufurd D, Hayden MR. A worldwide assessment of the frequency of suicide, suicide attempts, or psychiatric hospitalization after predictive testing for Huntington disease. *Am J Hum Genet.* 1999;64(5):1293–1304. <http://doi.org/10.1086/302374>.
41. Almqvist EW, Brinkman RR, Wiggins S, Hayden MR. Canadian Collaborative Study of Predictive Testing. Psychological consequences and predictors of adverse events in the first 5 years after predictive testing for Huntington's disease. *Clin Genet.* 2003;64(4):300–309. <http://doi.org/10.1034/j.1399-0004.2003.00157.x>.
42. Broadstock M, Michie S, Marteau T. Psychological consequences of predictive genetic testing: a systematic review. *Eur J Hum Genet.* 2000;8(10):731–738. <http://doi.org/10.1038/sj.ejhg.5200532>.
43. Gargiulo M, Lejeune S, Tanguy ML, et al. Long-term outcome of presymptomatic testing in Huntington disease. *Eur J Hum Genet.* 2009;17(2):165–171. <http://doi.org/10.1038/ejhg.2008.146>.
44. van der Meer LB, van Duijn E, Giltay EJ, Tibben A. Do attachment style and emotion regulation strategies indicate distress in predictive testing? *J Genet Couns.* 2015;24(5):862–871. <http://doi.org/10.1007/s10897-015-9822-z>.
45. Lêdo S, Ramires A, Leite Â, Dinis MAP, Sequeiros J. Long-term predictors for psychological outcome of pre-symptomatic testing for late-onset neurological diseases. *Eur J Med Genet.* 2018;61(10):575–580. <http://doi.org/10.1016/j.ejmg.2018.03.010>.
46. Green RC, Roberts JS, Cupples LA, et al. Disclosure of APOE genotype for risk of Alzheimer's disease. *N Engl J Med.* 2009;361(3):245–254. <http://doi.org/10.1056/NEJMoa0809578>.
47. Paulsen JS, Nance M, Kim JJ, et al. A review of quality of life after predictive testing for and earlier identification of neurodegenerative diseases. *Prog Neurobiol.* 2013;110:2–28. <http://doi.org/10.1016/j.pneurobio.2013.08.003>.
48. Winnberg E, Winnberg U, Pohlkamp L, Hagberg A. What to do with a second chance in life? Long-term experiences of non-carriers of Huntington's disease. *J Genet Couns.* 2018;27(6):1438–1446. <http://doi.org/10.1007/s10897-018-0257-1>.
49. Goldman JS. Genetic testing and counseling in the diagnosis and management of young-onset dementias. *Psychiatr Clin North Am.* 2015;38(2):295–308. <http://doi.org/10.1016/j.psc.2015.01.008>.
50. MacLeod R, Tibben A, Frontali M, et al. Recommendations for the predictive genetic test in Huntington's disease. *Clin Genet.* 2013;83(3):221–231. <http://doi.org/10.1111/j.1399-0004.2012.01900.x>.
51. Crook A, Williams K, Adams L, Blair I, Rowe DB. Predictive genetic testing for amyotrophic lateral sclerosis and frontotemporal dementia: genetic counselling considerations. *Amyotroph Lateral Scler Frontotemporal Degener.* 2017;18(7-8):475–485. <http://doi.org/10.1080/21678421.2017.1332079>.
52. Molinuevo JL, Pintor L, Peri JM, et al. Emotional reactions to predictive testing in Alzheimer's disease and other inherited dementias. *Am J Alzheimers Dis Other Demen.* 2005;20(4):233–238. <http://doi.org/10.1177/153331750502000408>.
53. Fanos JH, Gronka S, Wu J, Stanislaw C, Andersen PM, Benatar M. Impact of presymptomatic genetic testing for familial amyotrophic lateral sclerosis. *Genet Med.* 2011;13(4):342–348. <http://doi.org/10.1097/GIM.0b013e318204d004>.
54. Crozier S, Robertson N, Dale M. The psychological impact of predictive genetic testing for Huntington's disease: a systematic review of the literature. *J Genet Couns.* 2015;24(1):29–39. <http://doi.org/10.1007/s10897-014-9755-y>.
55. Shaw C, Abrams K, Marteau TM. Psychological impact of predicting individuals' risks of illness: a systematic review. *Soc Sci Med.* 1999;49(12):1571–1598. [http://doi.org/10.1016/s0277-9536\(99\)00244-0](http://doi.org/10.1016/s0277-9536(99)00244-0).
56. Heshka JT, Pallechi C, Howley H, Wilson B, Wells PS. A systematic review of perceived risks, psychological and behavioral impacts of genetic testing. *Genet Med.* 2008;10(1):19–32. <http://doi.org/10.1097/GIM.0b013e31815f524f>.
57. Goh AMY, Chiu E, Yastrubetskaya O, et al. Perception, experience, and response to genetic discrimination in Huntington's disease: the Australian results of the International RESPOND-HD study. *Genet Test Mol Biomarkers.* 2013;17(2):115–121. <http://doi.org/10.1089/gtmb.2012.0288>.
58. Dufrasne S, Roy M, Galvez M, Rosenblatt DS. Experience over fifteen years with a protocol for predictive testing for Huntington disease. *Mol Genet Metab.* 2011;102(4):494–504. <http://doi.org/10.1016/j.ymgme.2010.12.001>.
59. Gooding HC, Linnenbringer EL, Burack J, Roberts JS, Green RC, Biesecker BB. Genetic susceptibility testing for Alzheimer disease: motivation to obtain information and control as precursors to coping with increased risk. *Patient Educ Couns.* 2006;64(1-3):259–267. <http://doi.org/10.1016/j.pcc.2006.03.002>.
60. Schwartz M, Brandel JP, Babonneau ML, et al. Genetic testing in prion disease: psychological consequences of the decisions to know or not to know. *Front Genet.* 2019;10:895. <http://doi.org/10.3389/fgene.2019.00895>.
61. Owen J, Beck J, Campbell T, et al. Predictive testing for inherited prion disease: report of 22 years experience. *Eur J Hum Genet.* 2014;22(12):1351–1356. <http://doi.org/10.1038/ejhg.2014.42>.
62. Goldman JS. Predictive genetic counseling for neurodegenerative diseases: past, present, and future. *Cold Spring Harb Perspect Med.* 2020;10(7):a036525. <http://doi.org/10.1101/cshperspect.a036525>.
63. Crook A, Jacobs C, Newton-John T, O'Shea R, McEwen A. Genetic counseling and testing practices for late-onset neurodegenerative disease: a systematic review. *J Neurol.* 2022;269(2):676–692. <http://doi.org/10.1007/s00415-021-10461-5>.
64. MacLeod R, Beach A, Henriques S, Knopp J, Nelson K, Kerzin-Storarr L. Experiences of predictive testing in young people at risk of Huntington's disease, familial cardiomyopathy or hereditary breast and ovarian cancer. *Eur J Hum Genet.* 2014;22(3):396–401. <http://doi.org/10.1038/ejhg.2013.143>.
65. Uflacker A, Doraiswamy PM, Rechitsky S, See T, Geschwind M, Tur-Kaspa I. Preimplantation genetic diagnosis (PGD) for genetic prion disorder due to F198S mutation in the PRNP gene. *JAMA Neurol.* 2014;71(4):484–486. <http://doi.org/10.1001/jamaneurol.2013.5884>.
66. Vallabh SM, Minikel EV, Schreiber SL, Lander ES. Towards a treatment for genetic prion disease: trials and biomarkers. *Lancet Neurol.* 2020 1;19(4):361–368. [http://doi.org/10.1016/S1474-4422\(19\)30403-X](http://doi.org/10.1016/S1474-4422(19)30403-X).
67. Minikel EV, Zhao HT, Le J, et al. Prion protein lowering is a disease-modifying therapy across prion disease stages, strains and endpoints. *Nucleic Acids Res.* 2020 10;48(19):10615–10631. <http://doi.org/10.1093/nar/gkaa616>.

Supplemental Information

Table S1. Evidence for pathogenicity of reported PRNP variants. Four kinds of evidence are provided: high penetrance is supported by Mendelian segregation (at least three cases within a family) and confirmed *de novo* variants in cases, while increased risk is supported by enrichment in observed cases versus controls,¹ and observation of cases homozygous for the variant. According to ACMG/AMP guidelines, *de novo* protein-altering variants and case-control enrichment are considered strong and moderate evidence of pathogenicity, respectively.² While the guidelines generally designate Mendelian segregation as merely supportive evidence,² the knowledge that *PRNP* is the single causal gene in prion disease, with all known pathogenic variants occurring in the protein-coding region, raises the prior that a rare protein-coding variant that seen to segregate with prion disease is causal. Finally, a homozygous rare protein-coding variant in a case, particularly in the context of a negative family history, is suggestive of increased risk that cumulatively reaches clinical significance when two copies of the variant allele are present. Table adapted with permission from cureffi.org;³ which provides further discussion of evidence types.

*The four octapeptide repeats in human *PRNP* differ in codon choice at the DNA level while being identical at the protein level. Different DNA sequence variants giving rise to the same protein change have been reported in different families, and would have various distinct representations in HGVS nomenclature.⁴

Table S1. Evidence for pathogenicity of reported PRNP variants.

Variant	HGVS	Evidence for high penetrance	Evidence for increased risk	Refs	Comments
P39L	NM000311.5: p.Pro39Leu			4	
2-OPRD	*			5,6	
1-OPRI	*			7,8	
2-OPRI	*			9	
3-OPRI	*			10	
4-OPRI	*			11	most cases have negative family history
5-OPRI	*	Mendelian segregation		12	
6-OPRI	*	Mendelian segregation		13	
7-OPRI	*	Mendelian segregation		14	
8-OPRI	*	Mendelian segregation		14,15	
9-OPRI	*	Mendelian segregation, <i>de novo</i>		16,17	
12-OPRI	*	Mendelian segregation		18	

P84S	NM000311.5: p.Pro84Ser			19	
S97N	NM000311.5: p.Ser97Asn			20	
P102L	NM000311.5: p.Pro102Leu	Mendelian segregation	case/control enrichment	21	
P105L	NM000311.5: p.Pro105Leu	Mendelian segregation		22	2 sibs affected & genotyped, 1 ungenotyped parent likely affected
P105S	NM000311.5: p.Pro105Ser			23	
P105T	NM000311.5: p.Pro105Thr	Mendelian segregation		24	
G114V	NM000311.5: p.Gly114Val	Mendelian segregation		25,26	pedigree suggests high penetrance though not 100%
A117V	NM000311.5: p.Ala117Val	Mendelian segregation	case/control enrichment	27	
129insLGGLG GYV	NM000311.5: p.129insLGGLGGY V	<i>de novo</i>		28	
G131V	NM000311.5: p.Gly131Val			29,30	positive family history in one case
G131R	NM000311.5: p.Gly131Arg			31	positive family history
S132I	NM000311.5: p.Ser132Ile	Mendelian segregation		32	extensive family history, only proband genotyped
A133V	NM000311.5: p.Ala133Val			33	
R136S	NM000311.5: p.Arg136Ser		2 homozygotes	34	
Y145X	NM000311.5: p.Tyr145Ter			35	
R148H	NM000311.5: p.Arg148His			36	
R156C	NM000311.5: p.Arg156Cys			37	
Q160X	NM000311.5: p.Gln160Ter	Mendelian segregation		38	
Y162X	NM000311.5: p.Tyr162Ter	Mendelian segregation		39	
Y163X	NM000311.5: p.Tyr163Ter	Mendelian segregation		40,41	
D167G	NM000311.5: p.Asp167Gly			42	
D167N	NM000311.5: p.Asp167Asn			43	

Y169X	NM000311.5: p.Tyr169Ter	Mendelian segregation		41	
V176G	NM000311.5: p.Val176Gly			44	
D178Efs25X	NM000311.5: p.Asp178Glufs25T er	Mendelian segregation		45	only proband genotyped
D178N	NM000311.5: p.Asp178Asn	Mendelian segregation, <i>de novo</i>	case/control enrichment	46,47	
V180I	NM000311.5: p.Val180Ile		case/control enrichment	48	
T183A	NM000311.5: p.Thr183Ala	Mendelian segregation		49	
H187R	NM000311.5: p.His187Arg	Mendelian segregation		50	
T188A	NM000311.5: p.Thr188Ala			51	
T188K	NM000311.5: p.Thr188Lys			52	multiple cases with negative family history
T188R	NM000311.5: p.Thr188Arg			52,53	
V189I	NM000311.5: p.Val189Ile			54	
T193I	NM000311.5: p.Thr193Ile			55	
K194E	NM000311.5: p.Lys194Glu			56	
E196A	NM000311.5: p.Glu196Ala			57	
E196K	NM000311.5: p.Glu196Lys	Mendelian segregation		58	only proband genotyped
F198S	NM000311.5: p.Phe198Ser	Mendelian segregation		59,60	
F198V	NM000311.5: p.Phe198Val			20	
E200D	NM000311.5: p.Glu200Asp			61	
E200G	NM000311.5: p.Glu200Gly			62	
E200K	NM000311.5: p.Glu200Lys	Mendelian segregation	homozygote, case/control enrichment	27	
T201S	NM000311.5: p.Thr201Ser			63	
D202G	NM000311.5: p.Asp202Gly	Mendelian segregation		64	only proband genotyped
D202N	NM000311.5: p.Asp202Asn			65	
V203I	NM000311.5: p.Val203Ile		homozygote	66	
R208C	NM000311.5: p.Arg208Cys			20	
R208H	NM000311.5: p.Arg208His			67	

V210I	NM000311.5: p.Val210Ile		case/control enrichment	68,69	
E211D	NM000311.5: p.Glu211Asp	Mendelian segregation		70	supplement describes one family with 3 affected
E211Q	NM000311.5: p.Glu211Gln			58	2 sibs affected
Q212P	NM000311.5: p.Gln212Pro		homozygote	43	
I215V	NM000311.5: p.Ile215Val			71	
Q217R	NM000311.5: p.Gln217Arg			60	2 affected
Y218N	NM000311.5: p.Tyr218Asn	Mendelian segregation		72	
A224V	NM000311.5: p.Ala224Val			73	
Y225C	NM000311.5: p.Tyr225Cys			74	
Y226X	NM000311.5: p.Tyr226Ter			75	
Q227X	NM000311.5: p.Gln227Ter			75	
M232R	NM000311.5: p.Met232Arg			48	
M232T	NM000311.5: p.Met232Thr			76	
P238S	NM000311.5: p.Pro238Ser			77	

Supplemental references

1. Minikel EV, Vallabh SM, Lek M, Estrada K, Samocha KE, Sathirapongsasuti JF, McLean CY, Tung JY, Yu LPC, Gambetti P, Blevins J, Zhang S, Cohen Y, Chen W, Yamada M, Hamaguchi T, Sanjo N, Mizusawa H, Nakamura Y, Kitamoto T, Collins SJ, Boyd A, Will RG, Knight R, Ponto C, Zerr I, Kraus TFJ, Eigenbrod S, Giese A, Calero M, Pedro-Cuesta J de, Haik S, Laplanche JL, Bouaziz-Amar E, Brandel JP, Capellari S, Parchi P, Poleggi A, Ladogana A, O'Donnell-Luria AH, Karczewski KJ, Marshall JL, Boehnke M, Laakso M, Mohlke KL, Kähler A, Chambert K, McCarroll S, Sullivan PF, Hultman CM, Purcell SM, Sklar P, Lee SJ van der, Rozemuller A, Jansen C, Hofman A, Kraaij R, Rooij JGJ van, Ikram MA, Uitterlinden AG, Duijn CM van, (ExAC) EAC, Daly MJ, MacArthur DG. Quantifying prion disease penetrance using large population control cohorts. *Science Translational Medicine*. 2016 Jan 20;8(322):322ra9. PMID: 26791950
2. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, Grody WW, Hegde M, Lyon E, Spector E, Voelkerding K, Rehm HL, ACMG Laboratory Quality Assurance Committee. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med*. 2015 May;17(5):405–424. PMID: 25741868
3. Minikel E. Annotating the literature on pathogenicity of PRNP variants [Internet]. *cureffi.org*. 2017. Available from: <https://www.cureffi.org/2017/04/24/annotating-the-literature-on-pathogenicity-of-prnp-variants/>

4. Bernardi L, Cupidi C, Frangipane F, Anfossi M, Gallo M, Conidi ME, Vasso F, Colao R, Puccio G, Curcio SAM, Mirabelli M, Clodomiro A, Di Lorenzo R, Smirne N, Maletta R, Bruni AC. Novel N-terminal domain mutation in prion protein detected in 2 patients diagnosed with frontotemporal lobar degeneration syndrome. *Neurobiol Aging*. 2014 Nov;35(11):2657.e7-2657.e11. PMID: 25022973
5. Beck JA, Mead S, Campbell TA, Dickinson A, Wientjens DP, Croes EA, Van Duijn CM, Collinge J. Two-octapeptide repeat deletion of prion protein associated with rapidly progressive dementia. *Neurology*. 2001 Jul 24;57(2):354–356. PMID: 11468331
6. Capellari S, Parchi P, Wolff BD, Campbell J, Atkinson R, Posey DM, Petersen RB, Gambetti P. Creutzfeldt-Jakob disease associated with a deletion of two repeats in the prion protein gene. *Neurology*. 2002 Nov 26;59(10):1628–1630. PMID: 12451210
7. Laplanche JL, Delasnerie-Lauprêtre N, Brandel JP, Dussaucy M, Chatelain J, Launay JM. Two novel insertions in the prion protein gene in patients with late-onset dementia. *Hum Mol Genet*. 1995 Jun;4(6):1109–1111. PMID: 7655470
8. Pietrini V, Puoti G, Limido L, Rossi G, Di Fede G, Giaccone G, Mangieri M, Tedeschi F, Bondavalli A, Mancina D, Bugiani O, Tagliavini F. Creutzfeldt-Jakob disease with a novel extra-repeat insertional mutation in the PRNP gene. *Neurology*. 2003 Nov 11;61(9):1288–1291. PMID: 14610142
9. Hill AF, Joiner S, Beck JA, Campbell TA, Dickinson A, Poulter M, Wadsworth JDF, Collinge J. Distinct glycoform ratios of protease resistant prion protein associated with PRNP point mutations. *Brain*. 2006 Mar;129(Pt 3):676–685. PMID: 16415305
10. Nishida Y, Sodeyama N, Toru Y, Toru S, Kitamoto T, Mizusawa H. Creutzfeldt-Jakob disease with a novel insertion and codon 219 Lys/Lys polymorphism in PRNP. *Neurology*. 2004 Nov 23;63(10):1978–1979. PMID: 15557533
11. Kaski DN, Pennington C, Beck J, Poulter M, Uphill J, Bishop MT, Linehan JM, O'Malley C, Wadsworth JDF, Joiner S, Knight RSG, Ironside JW, Brandner S, Collinge J, Mead S. Inherited prion disease with 4-octapeptide repeat insertion: disease requires the interaction of multiple genetic risk factors. *Brain*. 2011 Jun;134(Pt 6):1829–1838. PMID: 21616973
12. Mead S, Webb TEF, Campbell TA, Beck J, Linehan JM, Rutherford S, Joiner S, Wadsworth JDF, Heckmann J, Wroe S, Doey L, King A, Collinge J. Inherited prion disease with 5-OPRI: phenotype modification by repeat length and codon 129. *Neurology*. 2007 Aug 21;69(8):730–738. PMID: 17709704
13. Mead S, Poulter M, Beck J, Webb TEF, Campbell TA, Linehan JM, Desbruslais M, Joiner S, Wadsworth JDF, King A, Lantos P, Collinge J. Inherited prion disease with six octapeptide repeat insertional mutation--molecular analysis of phenotypic heterogeneity. *Brain*. 2006 Sep;129(Pt 9):2297–2317. PMID: 16923955
14. Goldfarb LG, Brown P, McCombie WR, Goldgaber D, Swergold GD, Wills PR, Cervenakova L, Baron H, Gibbs CJ, Gajdusek DC. Transmissible familial Creutzfeldt-Jakob disease associated with five, seven, and eight extra octapeptide coding repeats in the PRNP gene. *Proc Natl Acad Sci USA*. 1991 Dec 1;88(23):10926–10930. PMID: 1730445
15. Laplanche JL, Hachimi KH, Durieux I, Thuillet P, Defebvre L, Delasnerie-Lauprêtre N, Peoc'h K, Foncin JF, Destée A. Prominent psychiatric features and early onset in an inherited prion disease with a new insertional mutation in the prion protein gene. *Brain*. 1999 Dec;122 (Pt 12):2375–2386. PMID: 10581230
16. Krasemann S, Zerr I, Weber T, Poser S, Kretschmar H, Hunsmann G, Bodemer W. Prion disease associated with a novel nine octapeptide repeat insertion in the PRNP gene. *Brain Res Mol Brain Res*. 1995 Dec 1;34(1):173–176. PMID: 8750875

17. Sánchez-Valle R, Aróstegui JI, Yagüe J, Rami L, Lladó A, Molinuevo JL. First demonstrated de novo insertion in the prion protein gene in a young patient with dementia. *J Neurol Neurosurg Psychiatry*. 2008 Jul;79(7):845–846. PMID: 18559465
18. Kumar N, Boeve BF, Boot BP, Orr CF, Duffy J, Woodruff BK, Nair AK, Ellison J, Kuntz K, Kantarci K, Jack CR, Westmoreland BF, Fields JA, Baker M, Rademakers R, Parisi JE, Dickson DW. Clinical characterization of a kindred with a novel 12-octapeptide repeat insertion in the prion protein gene. *Arch Neurol*. 2011 Sep;68(9):1165–1170. PMCID: PMC3326586
19. Jones M, Odunsi S, du Plessis D, Vincent A, Bishop M, Head MW, Ironside JW, Gow D. Gerstmann-Straüssler-Scheinker disease: novel PRNP mutation and VGKC-complex antibodies. *Neurology*. 2014 Jun 10;82(23):2107–2111. PMCID: PMC4118501
20. Zheng L, Longfei J, Jing Y, Xinqing Z, Haiqing S, Haiyan L, Fen W, Xiumin D, Jianping J. PRNP mutations in a series of apparently sporadic neurodegenerative dementias in China. *Am J Med Genet B Neuropsychiatr Genet*. 2008 Sep 5;147B(6):938–944. PMID: 18425766
21. Webb TEF, Poulter M, Beck J, Uphill J, Adamson G, Campbell T, Linehan J, Powell C, Brandner S, Pal S, Siddique D, Wadsworth JD, Joiner S, Alner K, Petersen C, Hampson S, Rhymes C, Treacy C, Storey E, Geschwind MD, Nemeth AH, Wroe S, Collinge J, Mead S. Phenotypic heterogeneity and genetic modification of P102L inherited prion disease in an international series. *Brain*. 2008 Oct;131(Pt 10):2632–2646. PMCID: PMC2570713
22. Yamada M, Itoh Y, Inaba A, Wada Y, Takashima M, Satoh S, Kamata T, Okeda R, Kayano T, Suematsu N, Kitamoto T, Otomo E, Matsushita M, Mizusawa H. An inherited prion disease with a PrP P105L mutation: clinicopathologic and PrP heterogeneity. *Neurology*. 1999 Jul 13;53(1):181–188. PMID: 10408557
23. Tunnell E, Wollman R, Mallik S, Cortes CJ, Dearmond SJ, Mastrianni JA. A novel PRNP-P105S mutation associated with atypical prion disease and a rare PrPSc conformation. *Neurology*. 2008 Oct 28;71(18):1431–1438. PMCID: PMC2676963
24. Rogaeva E, Zadikoff C, Ponsesse J, Schmitt-Ulms G, Kawarai T, Sato C, Salehi-Rad S, St George-Hyslop P, Lang AE. Childhood onset in familial prion disease with a novel mutation in the PRNP gene. *Arch Neurol*. 2006 Jul;63(7):1016–1021. PMID: 16831973
25. Rodriguez MM, Peoc'h K, Haïk S, Bouchet C, Vernengo L, Mañana G, Salamano R, Carrasco L, Lenne M, Beaudry P, Launay JM, Laplanche JL. A novel mutation (G114V) in the prion protein gene in a family with inherited prion disease. *Neurology*. 2005 Apr 26;64(8):1455–1457. PMID: 15851745
26. Liu Z, Jia L, Piao Y, Lu D, Wang F, Lv H, Lu Y, Jia J. Creutzfeldt-Jakob disease with PRNP G114V mutation in a Chinese family. *Acta Neurol Scand*. 2010 Jun;121(6):377–383. PMID: 20028338
27. Hsiao K, Meiner Z, Kahana E, Cass C, Kahana I, Avrahami D, Scarlato G, Abramsky O, Prusiner SB, Gabizon R. Mutation of the prion protein in Libyan Jews with Creutzfeldt-Jakob disease. *N Engl J Med*. 1991 Apr 18;324(16):1091–1097. PMID: 2008182
28. Hinnell C, Coulthart MB, Jansen GH, Cashman NR, Lauzon J, Clark A, Costello F, White C, Midha R, Wiebe S, Furtado S. Gerstmann-Straussler-Scheinker disease due to a novel prion protein gene mutation. *Neurology*. 2011 Feb 1;76(5):485–487. PMID: 21282596
29. Panegyres PK, Toufexis K, Kakulas BA, Cernevakova L, Brown P, Ghetti B, Piccardo P, Dlouhy SR. A new PRNP mutation (G131V) associated with Gerstmann-Straussler-Scheinker disease. *Arch Neurol*. 2001 Nov;58(11):1899–1902. PMID: 11709001

30. Jansen C, Parchi P, Capellari S, Strammiello R, Dopfer EGP, van Swieten JC, Kamphorst W, Rozemuller AJM. A second case of Gerstmann-Sträussler-Scheinker disease linked to the G131V mutation in the prion protein gene in a Dutch patient. *J Neuropathol Exp Neurol*. 2011 Aug;70(8):698–702. PMID: 21760536
31. Alshaikh JT, Qin K, Zhao L, Mastrianni JA. A novel PRNP-G131R variant associated with familial prion disease. *Neurology Genetics* [Internet]. Wolters Kluwer Health, Inc. on behalf of the American Academy of Neurology; 2020 Aug 1 [cited 2022 Apr 27];6(4). Available from: <https://ng.neurology.org/content/6/4/e454>
32. Hilton DA, Head MW, Singh VK, Bishop M, Ironside JW. Familial prion disease with a novel serine to isoleucine mutation at codon 132 of prion protein gene (PRNP). *Neuropathol Appl Neurobiol*. 2009 Feb;35(1):111–115. PMID: 19187063
33. Rowe DB, Lewis V, Needham M, Rodriguez M, Boyd A, McLean C, Roberts H, Masters CL, Collins SJ. Novel prion protein gene mutation presenting with subacute PSP-like syndrome. *Neurology*. 2007 Mar 13;68(11):868–870. PMID: 17353478
34. Ximelis T, Marín-Moreno A, Espinosa JC, Eraña H, Charco JM, Hernández I, Riveira C, Alcolea D, González-Roca E, Aldecoa I, Molina-Porcel L, Parchi P, Rossi M, Castilla J, Ruiz-García R, Gelpi E, Torres JM, Sánchez-Valle R. Homozygous R136S mutation in PRNP gene causes inherited early onset prion disease. *Alzheimer's Research & Therapy*. 2021 Oct 18;13(1):176.
35. Kitamoto T, Ohta M, Doh-ura K, Hitoshi S, Terao Y, Tateishi J. Novel missense variants of prion protein in Creutzfeldt-Jakob disease or Gerstmann-Sträussler syndrome. *Biochem Biophys Res Commun*. 1993 Mar 15;191(2):709–714. PMID: 8461023
36. Krebs B, Lederer RM, Windl O, Grasbon-Frodl EM, Zerr I, Kretzschmar HA. Creutzfeldt-Jakob disease associated with an R148H mutation of the prion protein gene. *Neurogenetics*. 2005 May;6(2):97–100. PMID: 15776279
37. Kenny J, Woollacott I, Koriath C, Hosszu L, Adamson G, Rudge P, Rossor MN, Collinge J, Rohrer JD, Mead S. A novel prion protein variant in a patient with semantic dementia. *J Neurol Neurosurg Psychiatry*. 2017 Oct;88(10):890–892. PMID: PMC5629930
38. Fong JC, Rojas JC, Bang J, Legati A, Rankin KP, Forner S, Miller ZA, Karydas AM, Coppola G, Grouse CK, Ralph J, Miller BL, Geschwind MD. Genetic Prion Disease Caused by PRNP Q160X Mutation Presenting with an Orbitofrontal Syndrome, Cyclic Diarrhea, and Peripheral Neuropathy. *J Alzheimers Dis*. 2017;55(1):249–258. PMID: PMC5149415
39. Bommarito G, Cellerino M, Prada V, Venturi C, Capellari S, Cortelli P, Mancardi GL, Parchi P, Schenone A. A novel prion protein gene-truncating mutation causing autonomic neuropathy and diarrhea. *Eur J Neurol*. 2018 Aug;25(8):e91–e92. PMID: 29984897
40. Mead S, Gandhi S, Beck J, Caine D, Gallujipali D, Carswell C, Hyare H, Joiner S, Ayling H, Lashley T, Linehan JM, Al-Doujaily H, Sharps B, Revesz T, Sandberg MK, Reilly MM, Koltzenburg M, Forbes A, Rudge P, Brandner S, Warren JD, Wadsworth JDF, Wood NW, Holton JL, Collinge J. A novel prion disease associated with diarrhea and autonomic neuropathy. *N Engl J Med*. 2013 Nov 14;369(20):1904–1914. PMID: PMC3863770
41. Capellari S, Baiardi S, Rinaldi R, Bartoletti-Stella A, Graziano C, Piras S, Calandra-Buonaura G, D'Angelo R, Terziotti C, Lodi R, Donadio V, Pironi L, Cortelli P, Parchi P. Two novel PRNP truncating mutations broaden the spectrum of prion amyloidosis. *Annals of Clinical and Translational Neurology*. 2018;5(6):777–783.

42. Bishop MT, Pennington C, Heath CA, Will RG, Knight RSG. PRNP variation in UK sporadic and variant Creutzfeldt Jakob disease highlights genetic risk factors and a novel non-synonymous polymorphism. *BMC Med Genet.* 2009 Dec 26;10:146. PMID: PMC2806268
43. Kroll F, Dimitriadis A, Campbell T, Darwent L, Collinge J, Mead S, Vire E. Prion protein gene mutation detection using long-read Nanopore sequencing. *Sci Rep.* 2022 May 18;12(1):8284. PMID: PMC9117325
44. Simpson M, Johanssen V, Boyd A, Klug G, Masters CL, Li QX, Pamphlett R, McLean C, Lewis V, Collins SJ. Unusual clinical and molecular-pathological profile of gerstmann-Sträussler-Scheinker disease associated with a novel PRNP mutation (V176G). *JAMA Neurol.* 2013 Sep 1;70(9):1180–1185. PMID: 23857164
45. Matsuzono K, Ikeda Y, Liu W, Kurata T, Deguchi S, Deguchi K, Abe K. A novel familial prion disease causing pan-autonomic-sensory neuropathy and cognitive impairment. *Eur J Neurol.* 2013 May;20(5):e67-69. PMID: 23577609
46. Medori R, Tritschler HJ, LeBlanc A, Villare F, Manetto V, Chen HY, Xue R, Leal S, Montagna P, Cortelli P. Fatal familial insomnia, a prion disease with a mutation at codon 178 of the prion protein gene. *N Engl J Med.* 1992 Feb 13;326(7):444–449. PMID: PMC6151859
47. Dagvadorj A, Petersen RB, Lee HS, Cervenakova L, Shatunov A, Budka H, Brown P, Gambetti P, Goldfarb LG. Spontaneous mutations in the prion protein gene causing transmissible spongiform encephalopathy. *Ann Neurol.* 2002 Sep;52(3):355–359. PMID: 12205650
48. Hitoshi S, Nagura H, Yamanouchi H, Kitamoto T. Double mutations at codon 180 and codon 232 of the PRNP gene in an apparently sporadic case of Creutzfeldt-Jakob disease. *J Neurol Sci.* 1993 Dec 15;120(2):208–212. PMID: 8138811
49. Nitrini R, Rosemberg S, Passos-Bueno MR, da Silva LS, Iughetti P, Papadopoulos M, Carrilho PM, Caramelli P, Albrecht S, Zatz M, LeBlanc A. Familial spongiform encephalopathy associated with a novel prion protein gene mutation. *Ann Neurol.* 1997 Aug;42(2):138–146. PMID: 9266722
50. Bütefisch CM, Gambetti P, Cervenakova L, Park KY, Hallett M, Goldfarb LG. Inherited prion encephalopathy associated with the novel PRNP H187R mutation: a clinical study. *Neurology.* 2000 Aug 22;55(4):517–522. PMID: 10953183
51. Collins S, Boyd A, Fletcher A, Byron K, Harper C, McLean CA, Masters CL. Novel prion protein gene mutation in an octogenarian with Creutzfeldt-Jakob disease. *Arch Neurol.* 2000 Jul;57(7):1058–1063. PMID: 10891990
52. Roeber S, Grasbon-Frodl EM, Windl O, Krebs B, Xiang W, Vollmert C, Illig T, Schröter A, Arzberger T, Weber P, Zerr I, Kretzschmar HA. Evidence for a pathogenic role of different mutations at codon 188 of PRNP. *PLoS One.* 2008 May 14;3(5):e2147. PMID: PMC2366066
53. Tartaglia MC, Thai JN, See T, Kuo A, Harbaugh R, Raudabaugh B, Cali I, Sattavat M, Sanchez H, DeArmond SJ, Geschwind MD. Pathologic evidence that the T188R mutation in PRNP is associated with prion disease. *J Neuropathol Exp Neurol.* 2010 Dec;69(12):1220–1227. PMID: PMC3136530
54. Di Fede G, Catania M, Atzori C, Moda F, Pasquali C, Indaco A, Grisoli M, Zuffi M, Guaita MC, Testi R, Taraglio S, Sessa M, Gusmaroli G, Spinelli M, Salzano G, Legname G, Tarletti R, Godi L, Pocchiari M, Tagliavini F, Imperiale D, Giaccone G. Clinical and neuropathological phenotype associated with the novel V189I mutation in the prion protein gene. *Acta Neuropathol Commun.* 2019 Jan 3;7(1):1. PMID: PMC6317215

55. Kotta K, Paspaltsis I, Bostantjopoulou S, Latsoudis H, Plaitakis A, Kazis D, Collinge J, Sklaviadis T. Novel mutation of the PRNP gene of a clinical CJD case. *BMC Infect Dis.* 2006 Nov 27;6:169. PMID: 1693557
56. Takada LT, Kim MO, Cleveland RW, Wong K, Forner SA, Gala H, Fong JC, Geschwind MD. Genetic prion disease: Experience of a rapidly progressive dementia center in the United States and a review of the literature. *Am J Med Genet B Neuropsychiatr Genet.* 2017 Jan;174(1):36–69. PMID: 2707989
57. Zhang H, Wang M, Wu L, Zhang H, Jin T, Wu J, Sun L. Novel prion protein gene mutation at codon 196 (E196A) in a septuagenarian with Creutzfeldt-Jakob disease. *J Clin Neurosci.* 2014 Jan;21(1):175–178. PMID: 23787189
58. Peoc'h K, Manivet P, Beaudry P, Attane F, Besson G, Hannequin D, Delasnerie-Lauprêtre N, Laplanche JL. Identification of three novel mutations (E196K, V203I, E211Q) in the prion protein gene (PRNP) in inherited prion diseases with Creutzfeldt-Jakob disease phenotype. *Hum Mutat.* 2000 May;15(5):482. PMID: 10790216
59. Dlouhy SR, Hsiao K, Farlow MR, Foroud T, Conneally PM, Johnson P, Prusiner SB, Hodes ME, Ghetti B. Linkage of the Indiana kindred of Gerstmann-Sträussler-Scheinker disease to the prion protein gene. *Nat Genet.* 1992 Apr;1(1):64–67. PMID: 1363809
60. Hsiao K, Dlouhy SR, Farlow MR, Cass C, Da Costa M, Conneally PM, Hodes ME, Ghetti B, Prusiner SB. Mutant prion proteins in Gerstmann-Sträussler-Scheinker disease with neurofibrillary tangles. *Nat Genet.* 1992 Apr;1(1):68–71. PMID: 1363810
61. Hassan A, Campbell T, Darwent L, Odd H, Green A, Collinge J, Mead S. Case report of homozygous E200D mutation of PRNP in apparently sporadic Creutzfeldt-Jakob disease. *BMC Neurol.* 2021 Jun 28;21(1):248. PMID: 3437416
62. Kim MO, Cali I, Oehler A, Fong JC, Wong K, See T, Katz JS, Gambetti P, Bettcher BM, Dearmond SJ, Geschwind MD. Genetic CJD with a novel E200G mutation in the prion protein gene and comparison with E200K mutation cases. *Acta Neuropathol Commun.* 2013 Dec 12;1:80. PMID: 2480091
63. Mok TH, Koriath C, Jaunmuktane Z, Campbell T, Joiner S, Wadsworth JDF, Hosszu LLP, Brandner S, Parvez A, Truelsén TC, Lund EL, Saha R, Collinge J, Mead S. Evaluating the causality of novel sequence variants in the prion protein gene by example. *Neurobiol Aging.* 2018 Nov;71:265.e1-265.e7. PMID: 3075539
64. Heinemann U, Krasnianski A, Meissner B, Grasbon-Frodl EM, Kretzschmar HA, Zerr I. Novel PRNP mutation in a patient with a slow progressive dementia syndrome. *Med Sci Monit.* 2008 May;14(5):CS41-43. PMID: 18443555
65. Piccardo P, Dlouhy SR, Lievens PM, Young K, Bird TD, Nochlin D, Dickson DW, Vinters HV, Zimmerman TR, Mackenzie IR, Kish SJ, Ang LC, De Carli C, Pocchiari M, Brown P, Gibbs CJ, Gajdusek DC, Bugiani O, Ironside J, Tagliavini F, Ghetti B. Phenotypic variability of Gerstmann-Sträussler-Scheinker disease is associated with prion protein heterogeneity. *J Neuropathol Exp Neurol.* 1998 Oct;57(10):979–988. PMID: 9786248
66. Komatsu J, Sakai K, Hamaguchi T, Sugiyama Y, Iwasa K, Yamada M. Creutzfeldt-Jakob disease associated with a V203I homozygous mutation in the prion protein gene. *Prion.* 2014;8(5):336–338. PMID: 24601383
67. Mastrianni JA, Iannicola C, Myers RM, DeArmond S, Prusiner SB. Mutation of the prion protein gene at codon 208 in familial Creutzfeldt-Jakob disease. *Neurology.* 1996 Nov;47(5):1305–1312. PMID: 8909447

68. Ripoll L, Laplanche JL, Salzmann M, Jouvét A, Planques B, Dussaucy M, Chatelain J, Beaudry P, Launay JM. A new point mutation in the prion protein gene at codon 210 in Creutzfeldt-Jakob disease. *Neurology*. 1993 Oct;43(10):1934–1938. PMID: 8105421
69. Pocchiari M, Salvatore M, Cutruzzolá F, Genuardi M, Allocatedelli CT, Masullo C, Macchi G, Alemá G, Galgani S, Xi YG. A new point mutation of the prion protein gene in Creutzfeldt-Jakob disease. *Ann Neurol*. 1993 Dec;34(6):802–807. PMID: 7902693
70. Peoc'h K, Levavasseur E, Delmont E, De Simone A, Laffont-Proust I, Privat N, Chebaro Y, Chapuis C, Bedoucha P, Brandel JP, Laquerriere A, Kemeny JL, Hauw JJ, Borg M, Rezaei H, Derreumaux P, Laplanche JL, Haïk S. Substitutions at residue 211 in the prion protein drive a switch between CJD and GSS syndrome, a new mechanism governing inherited neurodegenerative disorders. *Hum Mol Genet*. 2012 Dec 15;21(26):5417–5428. PMID: 22965875
71. Muñoz-Nieto M, Ramonet N, López-Gastón JI, Cuadrado-Corrales N, Calero O, Díaz-Hurtado M, Ipiens JR, Ramón y Cajal S, de Pedro-Cuesta J, Calero M. A novel mutation I215V in the PRNP gene associated with Creutzfeldt-Jakob and Alzheimer's diseases in three patients with divergent clinical phenotypes. *J Neurol*. 2013 Jan;260(1):77–84. PMID: 22763467
72. Alzualde A, Indakoetxea B, Ferrer I, Moreno F, Barandiaran M, Gorostidi A, Estanga A, Ruiz I, Calero M, van Leeuwen FW, Atares B, Juste R, Rodríguez-Martínez AB, López de Munain A. A novel PRNP Y218N mutation in Gerstmann-Sträussler-Scheinker disease with neurofibrillary degeneration. *J Neuropathol Exp Neurol*. 2010 Aug;69(8):789–800. PMID: 20613639
73. Watts JC, Giles K, Serban A, Patel S, Oehler A, Bhardwaj S, Guan S, Greicius MD, Miller BL, DeArmond SJ, Geschwind MD, Prusiner SB. Modulation of Creutzfeldt-Jakob disease prion propagation by the A224V mutation. *Ann Neurol*. 2015 Oct;78(4):540–553. PMCID: PMC4711268
74. Bagyinszky E, Yang Y, Giau VV, Youn YC, An SSA, Kim S. Novel prion mutation (p.Tyr225Cys) in a Korean patient with atypical Creutzfeldt-Jakob disease. *Clin Interv Aging*. 2019;14:1387–1397. PMCID: PMC6683949
75. Jansen C, Parchi P, Capellari S, Vermeij AJ, Corrado P, Baas F, Strammiello R, van Gool WA, van Swieten JC, Rozemuller AJM. Prion protein amyloidosis with divergent phenotype associated with two novel nonsense mutations in PRNP. *Acta Neuropathol*. 2010 Feb;119(2):189–197. PMCID: PMC2808512
76. Bratosiewicz J, Barcikowska M, Cervenakowa L, Brown P, Gajdusek DC, Liberski PP. A new point mutation of the PRNP gene in Gerstmann-Sträussler-Scheinker case in Poland. *Folia Neuropathol*. 2000;38(4):164–166. PMID: 11693719
77. Windl O, Giese A, Schulz-Schaeffer W, Zerr I, Skworc K, Arendt S, Oberdieck C, Bodemer M, Poser S, Kretzschmar HA. Molecular genetics of human prion diseases in Germany. *Hum Genet*. 1999 Sep;105(3):244–252. PMID: 10987652